Methylene-*gem*-Difluorocyclopropane Analogues of Nucleosides: Synthesis, Cyclopropene-Methylenecyclopropane Rearrangement, and Biological Activity¹

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Alkylation-elimination of adenine and 2-amino-6-chloropurine with *gem*-difluorocyclopropane dibromide **10** gave *E*- and *Z*-methylene-*gem*-difluorocyclopropanes **11a**, **11b**, **12a**, and **12b** and *gem*-difluorocyclopropenes **13a** and **13b**. Debenzylation of intermediates **11a**, **11b**, **12a**, and **12b** afforded *E*- and *Z*-methylenecyclopropanes **4a**, **4b**, **5a**, and **5b**. Hydrolysis of 2-amino-6-chloropurine derivatives **4b** and **5b** afforded guanine analogues **4c** and **5c**. Composition of products (except **14b**) obtained from alkylation-elimination reflects thermodynamically controlled cyclopropene-methylenecyclopropene rearrangement. The *E*-isomer **4a** was moderately active against human cytomegalovirus and along with the *Z*-isomer **5a** was active against leukemia L1210 and solid tumors in vitro.

Introduction

Recently, we described a series of methylenecyclopropane analogues of nucleosides 1 and 2 (Chart 1). The Z-isomers 1 are broad-spectrum antiviral agents especially effective against human cytomegalovirus (HC-MV).^{2,3} Replacing hydrogen with fluorine led in many cases to biologically active compounds.⁴ Among nucleosides fluorinated in the carbohydrate moiety,5 gemdifluoro analogues play a significant role. The anticancer drug gemcitabine⁶ (2'-deoxy-2',2'-difluorocytidine) and the acyclic nucleotide 9-(5,5-difluoro-5-phosphonopentyl)guanine, an inhibitor of purine nucleoside phosphorylase,⁷ are notable examples. More recently, we described⁸ also gem-difluorocyclopropane nucleoside analogues 3. In view of a potent antiviral activity of methylenecyclopropanes 1, structure-activity relationships in this series of analogues are under active investigation.^{9,10} As a part of this effort, gem-difluoro analogues 4 and 5 have become of interest.

Results and Discussion

The alkylation-elimination approach that was successfully applied for synthesis of nonfluorinated analogues^{2,3} **1** and **2** seemed the most promising for synthesis of **4** and **5**. The starting methylene-*gem*-difluorocyclopropane (**6**) was prepared from *gem*-difluorocyclopropane⁸ **7** by a modification of the described procedure¹¹ via selenide **8** and selenoxide **9** (Scheme 1). **6** was converted to the vicinal dibromocyclopropane **10** in 95% yield using pyridine·HBr₃ in CH₂Cl₂. Alkylation-elimination of sodium salt of adenine with **10** in DMF was perfomed at room temperature. A mixture of

Chart 1



Scheme 1



Series a: B = adenin-9-yl, series b: B = 2-amino-6-chloropurin-9-yl, series c: B = guanin-9-yl

 a Key: (a) (o-NO₂)C₆H₄SeCN, Bu₃P, THF; (b) H₂O₂, THF; (c) toluene, Δ ; (d) pyridine·HBr₃, CH₂Cl₂; (e) NaH, DMF; (f) BCl₃, CH₂Cl₂, -78 °C.

three products was obtained, the *E*,*Z*-isomers **11a** and **12a** (21 and 7%, respectively) and, surprisingly, *gem*difluorocyclopropene **13a** (31%). All three products were readily separated by chromatography on a silica gel column. Debenzylation of **11a** and **12a** using BCl₃ in CH₂Cl₂ at -78 °C was uneventful to give target analogues **4a** and **5a** in 75 and 77% yield, respectively. An

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Table 1. Antitumor Activity of Methylenecyclopropanes 1a, 4a, and 5a

^{*a*} Disk-diffusion assay,¹⁴ 500 μ g/disk, 200 units = 6 mm. ^{*b*} 11 μ g/disk.¹⁴

attempted deprotection of cyclopropene analogue **13a** led only to decomposition.

Alkylation–elimination of 2-amino-6-chloropurine with reagent **10** performed in a similar fashion (NaH in DMF at room temperature) gave a mixture of four compounds that were all separated by chromatography on silica gel. The yields of the *E*- and *Z*-isomers **11b** and **12b** and cyclopropene **13b** were 12, 3.5, and 17%. The product of a simple alkylation, bromo derivative **14b**, was also obtained in 10% yield. Debenzylation of **11b** and **12b** gave compounds **4b** and **5b** in 81 and 77% yield, respectively. Hydrolysis with 80% HCO₂H afforded guanine analogues **4c** (66%) and **5c** (71%).

The UV spectra of methylene–gem-difluorocyclopropanes reveal significant bathochromic shift throughout the entire region of spectrum when compared with nonfluorinated analogues.²

The *E*,*Z*-isomeric structures were assigned by NMR spectra. The H₈ values of the isomers with the heterocyclic base juxtaposed to hydroxymethyl group (Eisomers $4\mathbf{a} - \mathbf{c}$ and Z-isomers $1\mathbf{a} - \mathbf{c}$) are significantly deshielded relative to those of the Z-isomers 5a-c and the *E*-isomers $2\mathbf{a} - \mathbf{c}$. A selective deshielding of the $H_{1'}$ alkene proton by fluorine in the cis position was observed only in the *E*-isomers $4\mathbf{a}-\mathbf{c}$. The absence of alkene protons as well as the presence of three different methylene groups excluded structures 15a and 15b as possible alternatives for cyclopropenes 13a and 13b. These assignments were confirmed by the NOE experiments with isomers 4a and 5a. Thus, all protons of the cyclopropane moiety of E-isomer 4a located in the vicinity of H₈ exhibit NOE enhancements whereas none were found for a more distant $H_{1'}$. An opposite situation was encountered in the Z-isomer 5a. In this case, all cyclopropane protons are closer to $H_{1'}$ than to H_{8} , and consequently, NOE was observed at $H_{1'}$ but not at H_8 .

Because the *E*- and *Z*-isomers **4a** and **4c** and **5a** and 5c were the main targets of our study, we investigated a possibility of isomerization of cyclopropene **13b** into the corresponding methylenecyclopropanes. Base-catalyzed cyclopropene-methylene-cyclopropane rearrangements have been described in both nonfluorinated¹² and gem-difluorocyclopropane series.¹³ Isomerization of cyclopropene 13b using DBU in DMF at 0 °C (2 min) gave *E*-isomer **11b** (37%), *Z*-isomer **12b** (11.5%), and starting material **13b** (39%) that were separated by chromatography on a silica gel column. The ratio of these components resembled that obtained by alkylation-elimination of nucleic acid bases (Scheme 1). The result indicated that isomerization of cyclopropene 13b to the *E*- and *Z*-isomers **11b** and **12b** is possible but only to a limited extent.

Interconversion of all three components **11b**, **12b**, and **13b** (catalysis with DBU in DMF at room temperature) was investigated by HPLC. After 1 min, the average

ratio 11b:12b:13b = 3.6:1:5.4 resembled that found in the above-mentioned preparative experiment with cyclopropene 13b. This ratio was not changed after a prolonged reaction time or when excess of DBU was used, but decomposition of the products gradually occurred. Thus, the observed composition of products is thermodynamically controlled.

The shelf life of methylene–gem-difluorocyclopropanes $4\mathbf{a}-\mathbf{c}$ and $5\mathbf{a}-\mathbf{c}$ is limited, and a prolonged storage at low temperatures is mandatory. The least stable are the guanine analogues $4\mathbf{c}$ and $5\mathbf{c}$ where satisfactory elemental analyses could not be obtained although both compounds were fully characterized by spectroscopic methods.

Biological Activity

Analogues 4a, 4c, 5a, and 5c were tested against HCMV, HSV-1, HSV-2, EBV, HBV, VZV, and HIV-1 in previously described assays.² Only the E-isomer 4a had activity against the Towne strain of HCMV propagated in human foreskin fibroblast (HFF) cells (EC₅₀ 21 µM), and it was non-cytotoxic (CC₅₀ > 100 μ M). More moderate effects were observed against herpes simplex virus type 1 (HSV-1, EC₅₀ 70 μ M) in BSC-1 cells as determined by ELISA assay and EBV in Daudi cells (EC₅₀ 77 μ M). A decreased antiviral potency relative to that of nonfluorinated analogue² 1a can be explained by a limited capability of phosphorylation inside the infected cells. Lesser chemical stability of 4a may also be a factor. 4a and 5a also exhibited antitumor activity (Table 1). The E-isomer 4a was more selective than Z-isomer 5a. The activity of 4a corresponded to that of the nonfluorinated analogue 1a, but unlike 5a, E-isomer 2a was inactive. In contrast to 1a and 2a, compounds 4a and 5a were not substrates for adenosine deaminase.

Experimental Section

General Methods. See ref 2. The UV spectra were recorded in ethanol. The NMR spectra were determined at 300 or 400 MHz (¹H), 75 or 100 MHz (¹³C), and 376 MHz (¹⁹F) in DMSO d_6 unless specified otherwise. CFCl₃ was used as a reference for ¹⁹F spectra. For FAB-MS, the thioglycerol matrix was used.

2-Benzyloxymethyl-3,3-difluoro-1-methylenecyclopropane (6). The described procedure¹¹ starting from *gem*difluorocyclopropane⁸ **7** was modified as follows. Selenide **8** (4.4 g, 10.4 mmol) was treated with 30% H_2O_2 (10 mL) in THF (50 mL) at 0 °C. The reaction mixture was allowed to stand at room temperature for 16 h, and then it was diluted with EtOAc (100 mL) and washed with saturated $Na_2S_2O_3$ solution and water. The organic phase was dried over Na_2SO_4 , and the solvent was evaporated to give a crude selenoxide **9** as a yellow solid. A solution of **9** in toluene (50 mL) was heated at 80 °C for 48 h, and the solvent was evaporated. The residue was chromatographed on a silica gel column using hexane–EtOAc (97:3) to give product **6** (1.56 g, 71%) as a colorless oil. The ¹H, ¹³C, and ¹⁹F NMR spectral data corresponded to those reported.¹¹ (*E,Z*)-2-Benzyloxymethyl-1-bromo-1-bromomethyl-3,3difluorocyclopropane (10). A solution of **6** (1.05 g, 5.0 mmol) in CH₂Cl₂ (50 mL) was treated with pyridine hydrobromide perbromide (2.56 g, 8 mmol) at 0 °C. The reaction mixture was allowed to stand at room temperature for 16 h. It was diluted with EtOAc (100 mL) and washed with saturated Na₂S₂O₃-NaHCO₃ solution and water. The organic phase was dried over Na₂SO₄, and the solvent was evaporated to give a pale yellow liquid **10** (1.76 g, 95.1%), which was used directly in the next step. ¹H NMR (CDCl₃) (δ): 1.96–2.03 and 2.42–2.49 (m, 1H), 3.61–3.85 (m, 4H), 4.50–4.58 (m, 2H), 7.30–7.39 (m, 5H).

(*E*)-9-{[2-(Benzyloxymethyl)-3,3-difluorocyclopropylidene]methyl}adenine (11a), (*Z*)-9-{[2-(Benzyloxymethyl)-3,3-difluorocyclopropylidene]methyl}adenine (12a), and 9-{[2-(Benzyloxymethyl)-3,3-difluorocyclopropenyl]methyl}adenine (13a). A mixture of adenine (1.485 g, 11 mmol) and NaH (60% oil suspension, 600 mg, 12.5 mmol) in DMF (100 mL) was stirred at room temperature for 4 h. The mixture was cooled to 0 °C, a solution of dibromide 10 (1.76 g, 4.76 mmol) in DMF (5 mL) was added dropwise, and the stirring was continued at room temperature for 16 h. The solvent was evaporated in vacuo, and the residue was chromatographed on a silica gel column (CH₂Cl₂-MeOH, 49: 1) to give the products 11a (350 mg, 21.4%), 12a (120 mg, 7.4%), and 13a (510 mg, 31.2%) as white solids.

11a: mp 254–255 °C; UV max 284 nm (ϵ 6,000), 245 (ϵ 24,-100). ¹H NMR (δ): 3.23–3.28 (1H, m), 3.60 (1H, dd, J = 9.3 and 9.0 Hz), 3.85–3.91 (1H, m), 4.49 (1H, d, J = 12.3 Hz), 4.54 (1H, d, J = 12.3 Hz), 7.24–7.32 (5H, m), 7.50 (2H, s), 8.20 (1H, s), 8.25 (1H, d, J = 3.0 Hz), 8.58 (s, 1H). ¹³C NMR: 31.0 (t, J = 12.0 Hz), 66.5, 72.7, 107.5 (t, J = 7.1 Hz), 108.0 (t, J = 285.3 Hz), 119.2, 119.4, 128.1, 128.2, 128.9, 138.5, 139.0, 149.3, 154.3, 156.8. ¹⁹F NMR: 65.5 (d, J = 174.1 Hz), 77.8 (dd, J = 174.9, 6.0 Hz). EI-MS: 343 (M, 0.7), 342 (M – H, 1.1), 91 (100.0). HRMS calcd. for C₁₇H₁₄F₂N₅O (M – H) 342.1166, found: 342.1164. Anal. (C₁₇H₁₅F₂N₅O) C, H, N.

12a: mp 232–234 °C; UV max 284 nm (ϵ 6,700), 245 (ϵ 24,-200). ¹H NMR (δ): 3.02–3.07 (1H, m), 3.52 (1H, dd, J= 10.5, 9.6 Hz), 3.71–3.78 (1H, m), 4.54 (2H, s) 7.26–7.35 (5H, m), 7.48 (2H, s), 7.70 (1H, d, J= 2.1 Hz), 8.19 (1H, s), 8.23 (1H, s). ¹³C NMR: 28.4 (t, J= 12.1 Hz), 66.2, 72.4, 106.7 (t, J= 247.0 Hz), 107.6 (t, J= 7.8 Hz), 118.3, 119.4, 128.3, 129.0, 138.7, 139.0, 149.1, 154.5, 156.8. ¹⁹F NMR: 66.8 (d, J= 172.6 Hz), 79.2 (d, J= 173.7 Hz). EI-MS: 343 (M, 0.5), 342 (M – H, 1.0), 91 (100.0). HRMS calcd. for C₁₇H₁₄FN₅O (M – HF) 323.1182, found: 323.1179. Anal. (C₁₇H₁₅F₂N₅O) C, H, N, F.

13a: mp 195–196 °C; UV max 261 nm (ϵ 12,500), 209 (ϵ 22,500). ¹H NMR (δ): 4.30 (2H, s), 4.33 (2H, s), 5.45 (2H, s), 7.12–7.14 (2H, m), 7.24–7.32 (3H, m), 7.37 (2H, s), 8.17 (1H, s), 8.21 (1H, s). ¹³C NMR: 37.6, 61.7, 72.5, 103.6 (t, J = 269.3 Hz), 119.3, 127.3 (t, J = 11.0 Hz), 128.3, 128.4, 129.0, 129.2 (t, J = 11.0 Hz), 137.7, 141.2, 150.2, 153.5, 156.7. ¹⁹F NMR: 99.7. FAB-MS: 344 (M + H, 100.0). Anal. (C₁₇H₁₅F₂N₅O) C, H, N, F.

(E)-9-{[2-(Hydroxymethyl)-3,3-difluorocyclopropyli**denelmethyl**}**adenine (4a)**. Boron trichloride (1 M in CH₂-Cl₂, 5.8 mL, 5.8 mmol) was added to a solution of **11a** (200 mg, 0.58 mmol) in CH₂Cl₂ (50 mL) at -78 °C under N₂ over 10 min with stirring. The stirring was continued for 5 h at -78 °C. The reaction was quenched by a cautious addition of MeOH (10 mL), NaHCO₃ (5 g) was then added, and the mixture was stirred at room temperature for 4 h. The insoluble portion was filtered off, and it was washed with CH₂Cl₂-MeOH (4:1, 100 mL). The combined organic phase was evaporated, and the residue was chromatographed on a silica gel column (CH₂Cl₂–MeOH, 20:1) to give a white solid **4a** (110 mg, 74.8%), mp 268–270 °C; UV max 283 nm (ϵ 6,800), 243 (ϵ 27,300), 214 (ε 18,000). ¹H NMR (δ): 3.05-3.15 (1H, m), 3.62-3.68 (1H, m), 3.71-3.78 (1H, m), 5.39 (1H, t, J = 5.6 Hz), 7.48 (2H, s), 8.19 (1H, d, J = 2.4 Hz), 8.21 (1H, s), 8.68 (1H, s). ¹³C NMR: 33.4 (t, J = 11.0 Hz), 58.2, 108.2 (t, J = 285.4 Hz), 108.3 (t, J = 7.0 Hz), 118.6, 119.2, 138.9, 149.3, 154.3, 156.8. ¹⁹F NMR: 64.7 (d, *J* = 170.7 Hz), 78.2 (dd, *J* = 170.7 and 7.5 Hz). EI-MS: 253 (M, 41.9), 252 (M - H, 10.9), 236 (M - OH, 100.0).

HRMS calcd. for $C_{10}H_9F_2N_5O$ 253.0775, found: 253.0781. Anal. ($C_{10}H_9F_2N_5O$) C, H, N.

(Z)-9-{[2-(Hydroxymethyl)-3,3-difluorocyclopropylidene]methyl}adenine (5a). The reaction was performed as described for 4a (60 mg, 0.17 mmol of 12a) to give a white solid 5a (33 mg, 76.7%), mp 256–260 °C; UV max 283 nm (ϵ 5,700), 243 (ϵ 22,200), 214 (ϵ 14,100). ¹H NMR: 2.82–2.90 (1H, m), 3.44–3.53 (1H, m), 3.63–3.70 (1H, m), 5.24 (1H, brs), 7.48 (2H, s), 7.64 (1H, d, J = 2.0 Hz), 8.16 (1H, s), 8.22 (1H, s). ¹³C NMR: 31.2 (t, J = 11.0 Hz), 58.2, 107.1 (t, J = 286.4 Hz), 108.4 (t, J = 6.0 Hz), 117.5, 119.4, 138.8, 149.1, 154.5, 156.8. ¹⁹F NMR: 65.4 (d, J = 175.2 Hz), 79.3 (dd, J = 175.4, 6.3 Hz). EI-MS 253 (M, 12.3), 252 (M – H, 6.7), 236 (M – OH, 100.0). HRMS calcd. for C₁₀H₉F₂N₅O 253.0775, found: 253.0781. Anal. (C₁₀H₉F₂N₅O) C, H, N.

(*E*)-2-Amino-9-{[2-(benzyloxymethyl)-3,3 difluorocyclopropylidene]methyl}-6-chloropurine (11b), (*Z*)-2-Amino-9-{[2-(benzyloxymethyl)-3,3-difluorocyclopropylidene]methyl}-6-chloropurine (12b), 2-Amino-9-{[2-(benzyloxymethyl)-3,3-difluorocyclopropenyl]methyl}-6-chloropurine (13b), and (*E,Z*)-2-Amino-9-{[1-bromo-2-(benzyloxymethyl)-3,3-difluorocyclopropyl]methyl}-6-chloropurine (14b). The procedure described for adenine analogues was followed with 2-amino-6-chloropurine (4.25 g, 25 mmol), NaH (1.08 g, 22.5 mmol) in DMF (200 mL), and dibromide 10 (2.81 g, 8.00 mmol). The products were separated by column chromatography on silica gel using hexane–EtOAc (3:1) to give products 11b (350 mg, 11.6%), 12b (105 mg, 3.5%), 14b (350 mg, 9.6%), and 13b (510 mg, 16.9%).

11b: white solid, mp 138–139 °C; UV max 308 nm (ϵ 7,-800), 285 (ϵ 8,100), 240 (ϵ 26,000), 205 (ϵ 21,500). ¹H NMR (CDCl₃ δ): 2.85–2.96 (1H, m), 3.51–3.58 (1H, m), 3.88 (1H, ddd, J = 9.6, 6.0 and 1.8 Hz), 4.57 (2H, AB, J = 12.9 and 12.9 Hz), 5.36 (2H, s, 7.27–7.36 (5H, m), 7.91 (1H, d, J = 3.0 Hz), 8.78 (1H, s). ¹³C NMR: 30.4 (t, J = 12.0 Hz), 66.0, 73.7, 105.8 (t, J = 285.3 Hz), 108.5 (t, J = 8.1 Hz), 117.1, 125.2, 128.2, 128.4, 128.8, 137.2, 139.8, 152.0, 153.0, 159.8. ¹⁹F NMR: 60.6 (dd, J = 175.6 and 3.0 Hz), 72.2 (dd, J = 175.4 and 9.4 Hz). EI-MS: 379 (M, 1.2), 377 (M, 3.1), 91 (100.0). HRMS calcd. for C₁₇H₁₄³⁵ClF₂N₅O 377.0855, found: 377.0856. Anal. (C₁₇H₁₄-ClF₂N₅O) C, H, N, Cl, F.

12b: white solid, mp 120–122 °C; UV max 307 nm (ϵ 7,-900), 284 (ϵ 7,700), 241 (ϵ 24,700), 206 (ϵ 22,900). ¹H NMR (CDCl₃ δ): 2.85–2.92 (1H, m), 3.68 (2H, d, J = 7.2 Hz), 4.54 (1H, d, J = 11.7 Hz), 4.60 (1H, d, J = 12.3 Hz), 5.64 (2H, s), 7.27–7.35 (6H, m), 7.97 (1H, s). ¹³C NMR: 29.0 (t, J = 12.0 Hz), 65.8, 73.2, 105.6 (t, J = 288.4 Hz), 108.7 (t, J = 7.1 Hz), 116.0, 125.2, 128.0, 128.2, 128.8, 137.7, 139.2, 152.1, 152.5, 160.1. ¹⁹F NMR: 60.1 (d, J = 178.6 Hz), 72.2 (dd, J = 176.9 and 7.5 Hz). EI-MS: 379 (M, 0.7), 377 (M, 1.6), 91 (100.0). HRMS calcd. for C₁₇H₁₄³⁵ClF₂N₅O 377.0855, found: 377.0851.

13b: white solid, mp 133–134 °C; UV max 310 nm (ϵ 8,-000), 247 (ϵ 7,400), 220 (ϵ 28,200). ¹H NMR: 4.26 (2H, s), 4.33 (2H, s), 5.34 (2H, s), 7.00 (2H, s), 7.08–7.10 (2H, m), 7.24–7.32 (3H, m), 8.17 (1H, s). ¹³C NMR: 37.8, 61.8, 72.5, 103.5 (t, J= 269.3 Hz), 123.9, 127.0 (t, J= 12.0 Hz), 128.1, 128.4, 129.0, 129.4 (t, J = 12.0 Hz), 137.7, 143.4, 150.2, 154.7, 160.6. ¹⁹F NMR: 99.9. EI-MS: 379 (M, 1.5), 377 (M, 4.0), 91 (100.0). HRMS calcd. for C₁₇H₁₄³⁵ClF₂N₅O 377.0855, found: 377.0851.

14b: pale yellow solid, mp 51–54 °C; UV max 310 nm (ϵ 7,900), 248 (ϵ 7,300), 221 (ϵ 27,200). ¹H NMR (CDCl₃ δ): 2.57–2.65 (1H, m), 3.50–3.55 (1H, m), 3.68 (1H, dd, J = 11.4 and 6.4 Hz), 4.37 (1H, d, J = 12.0 Hz), 4.42 (1H, d, J = 12.4 Hz), 4.45 (1H, d, J = 15.2 Hz), 4.63 (1H, d, J = 15.6 Hz), 5.40 (brs, 2H), 7.15–7.17 (2H, m), 7.24–7.32 (3H, m), 7.95 (1H, s). ¹³C NMR: 32.9 (t, J = 7.8 Hz), 39.3 (t, J = 8.4 Hz), 48.6, 65.9, 73.1, 110.2 (dd, J = 222.0 and 217.0 Hz), 125.0, 127.7, 128.2, 128.7, 137.4, 142.3, 151.7, 154.2, 159.5. ¹⁹F NMR: 61.1 (d, J = 161.7 Hz), 64.9 (dd, J = 160.0 and 15.0 Hz). EI-MS: 459 (M, 2.7), 457 (M, 2.1), 91 (100.0). HRMS calcd. for C₁₇H₁₅-⁷⁹Br³⁵ClF₂N₅O 457.0117, found: 457.0116.

(E)-2-Amino-9-{[2-(hydroxymethyl)-3,3-difluorocyclopropylidene]methyl}-6-chloropurine (4b). This reaction was performed as described for adenine analogue 4a with 11b (360 mg, 0.95 mmol). The crude product was purified by silica gel column chromatography (CH₂Cl₂–MeOH, 30:1) to give a white solid **4b** (220 mg, 80.7%), mp 239–241 °C; UV max 309 nm (ϵ 7,000), 284 (ϵ 7,300), 240 (ϵ 24,000), 204 (ϵ 13,800). ¹H NMR (δ): 3.06–3.13 (1H, m), 3.58–3.64 (1H, m), 3.66–3.72 (1H, m), 5.30 (1H, t, J = 5.6 Hz), 7.14 (2H, s), 8.00 (1H, d, J = 2.4 Hz), 8.59 (1H, s). ¹³C NMR: 33.8 (t, J = 11.9 Hz), 58.3, 108.3 (t, J = 284.8 Hz), 109.2 (t, J = 8.0 Hz), 118.2, 123.9, 140.8, 150.6, 153.6, 161.0. ¹⁹F NMR: 64.7 (d, J = 171.4 and 7.5 Hz). EI-MS: 289 (M, 16.9), 287 (M, 52.3), 270 (M – OH, 100.0). HRMS calcd. for C₁₀H₈- ³⁵ClF₂N₅O 287.0385, found: 287.0383. Anal. (C₁₀H₈ClF₂N₅O) C, H, N, Cl, F.

(Z)-2-Amino-9-{[2-(hydroxymethyl)-3,3-difluorocyclopropylidene]methyl}-6-chloropurine (5b). The procedure described above was performed with 12b (130 mg, 0.35 mmol). The crude product was purified by chromatography (CH₂Cl₂-MeOH, 20:1) to give a white solid 5b (77 mg, 76.7%), mp 227–228 °C; UV max 307 nm (ϵ 7,700), 284 (ϵ 7,400), 240 (ϵ 24,500), 202 (ϵ 13,200). ¹H NMR (δ): 2.85–2.93 (1H, m), 3.48–3.55 (1H, m), 3.62–3.69 (1H, m), 5.23 (1H, brs), 7.07 (2H, s), 7.50 (1H, s), 8.06 (1H, s). ¹³C NMR: 31.4 (t, J = 11.9 Hz), 58.1, 106.9 (t, J = 287.0 Hz), 108.8 (t, J = 7.3 Hz), 116.8, 123.9, 140.1, 150.7, 153.2, 161.1. ¹⁹F NMR: 65.1 (d, J = 175.2 Hz), 79.0 (d, J = 175.2 Hz). EI-MS: 289 (M, 9.6), 287 (M, 28.5), 270 (M – OH, 100.0). HRMS calcd. for C₁₀H₈³⁵ClF₂N₅O 287.0385, found: 287.0380. Anal. (C₁₀H₈ClF₂N₅O) C, H, N, Cl, F.

(E)-9-{[2-(Hydroxymethyl)-3,3-difluorocyclopropylidene]methyl}guanine (4c). A solution of 4b (210 mg, 0.73 mmol) in formic acid (80%, 30 mL) was heated at 90 °C for 6 h. After cooling, the solution was evaporated, and the residue was dissolved in NH₃/MeOH (20%, 10 mL). The mixture was stirred at room temperature for 30 min. The volatile components were evaporated, and the crude product was crystallized from MeOH- $\hat{H_2}O$ (1:2, active carbon) to afford a white solid 4c (180 mg, 92%), mp 304–307 °C (decomp.); UV max 292 nm (\$\epsilon 5,500), 272 (\$\epsilon 7,500), 243 (\$\epsilon 28,100), 213 (\$\epsilon 16,000). ¹H NMR (δ): 3.01–3.09 (1H, m), 3.60–3.70 (2H, m), 5.31 (1H, t, J =5.6 Hz), 6.64 (2H, s), 7.86 (1H, d, J = 2.4 Hz), 8.27 (1H, s), 10.83 (1H, s). ¹³C NMR: 33.2 (t, J = 11.9 Hz), 58.1, 108.1 (t, *J* = 6.6 Hz), 108.2 (t, *J* = 285.7 Hz), 117.1, 118.2, 135.1, 151.1, 155.0, 157.2. ¹⁹F NMR: 64.9 (d, J = 170.7 Hz), 78.2 (dd, J =171.5 and 9.0 Hz). FAB-MS: (+ KCl) 308 (M + K, 6.9), 270 (M + H, 14.8), 253 (M + H - OH, 100.0).

(Z)-9-{[2-(Hydroxymethyl)-3,3-difluorocyclopropylidene]methyl}guanine (5c). The procedure described above was performed with 5b (60 mg, 0.21 mmol) to give a white solid 5c (40 mg, 71%), mp >350 °C (decomp.); UV max 292 nm (ϵ 5,400), 272 (ϵ 8,000), 244 (ϵ 28,400), 211 (ϵ 19,300). ¹H NMR (δ): 2.84–2.93 (1H, m), 3.47–3.53 (1H, m), 3.60–3.68 (1H, m), 5.20 (1H, t, J = 5.6 Hz), 6.60 (2H, s), 7.38 (1H, s), 7.64 (1H, s), 10.83 (1H, s). ¹³C NMR: 31.5 (t, J = 11.9 Hz), 58.1, 107.2 (t, J = 287.0 Hz), 107.8 (t, J = 6.4 Hz), 117.0, 117.3, 134.0, 150.8, 155.1, 157.2. ¹⁹F NMR: 64.1 (d, J = 172.6 Hz), 78.1 (dd, J = 174.1 and 7.9 Hz). FAB-MS: 270 (M + H, 67.7), 269 (M, 16.3), 232 (100.0).

Conversion of 13b to 11b and 12b. DBU (20 μ L, 0.13 mmol) was added to a solution of **13b** (410 mg, 1.09 mmol) in DMF (5 mL) at 0 °C. After 2 min the solvent was evaporated, and the residue was chromatographed on a silica gel column to give products **11b** (150 mg, 36.6%) and **12b** (47 mg, 11.5%) in addition to starting material **13b** (160 mg, 39%).

Cyclopropene–Methylenecyclopropane Rearrangement of 11b, 12b, and 13b. A solution of **11b, 12b,** or **13b** (0.24 μ M) in DMF (50 μ L) was treated with DBU (0.03 μ M) in DMF (5 μ L) at room temperature. After 1 min, a sample was analyzed by a reverse-phase HPLC (Waters μ Bondapak C₁₈, 300 \times 3.9 mm column, 45% MeCN in water, flow rate 0.9 mL/ min, detection at 310 nm). The average ratio **11b:12b:13b** from these experiments was 3.6:1:5.4. An excess of DBU or extension of the reaction time did not affect this ratio, but gradual decomposition was observed.

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Supporting Information Available: Comparison of NMR chemical shifts of fluorinated and nonfluorinated methylenecyclopropanes (Table 1), NOE data (Table 2), HPLC (cyclopropene-methylenecyclopropane rearrangement, Table 3), and UV spectra of **4a** and **1a** (Figure 1). This material is available free of charge via the Internet at http://pubs.acs.org.

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